NC COOPERATIVE

FECAL EGG COUNTING (FEC)

Is useful to determine:

- Which drugs are effective on that farm
- Which horses tend to always have low FECs (require less deworming)
- Which horses tend to always have high FECs (require more deworming)
- The interval between treatments that is required for the different drugs (this will be different for each drug)
- Are FECs getting lower over time? This could indicate that worms are being controlled better than when the program was started
- After having positive results for a few years, are FECs starting to rise? (this could indicate new resistance is developing)

What level shedder is my horse?

- FEC less than 200 = low shedder
- FEC between 200-500 = moderate shedder
- FEC greater than 500 = high shedder

Why is FEC Reduction Testing important to a strategic deworming plan?

FECRT is the only method currently available for detecting anthelmintic resistance in parasites of horses.

Fecal Egg Count Reduction (FECR) Calculation for Strongyles				
Horse I.D.	Pretreatment EPG	Post-treatment EPG ¹	Calculated FECR ²	
Buster	400	50	87.5%	
Fred	600	25	95.8%	
Mindy	150	0	100.0%	
¹ Optimally from a sample collected 10 to 14 days post-treatment		ent Herd Average	Herd Average 94.4% ³	

² Calculated as: (PreTx EPG) – (Post-Tx EPG)

PreTx EPG

³ Interpretation: >90% = effective anthelmintic

80 to 90% = equivocal; repeat in the future

<80% = worms are resistant; never use this product on this farm against strongyles

Source: https://aaep.org/horsehealth/internal-parasites-strategies-effective-parasite-control

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Supplies Needed:

- 2 small plastic cups
- Popsicle stick
- Kitchen scale
- 30cc Syringe
- Feca-Med Solution
- Tea strainer
- Pipettes
- Latex gloves (optional)

Steps:

- 1. Place 2 plastic cups on scale and zero
- 2. Add manure to reach 4g sample
- 3. Add 26cc Feca-Med Solution (shake well first!)
- 4. Mix solution and manure well squish on sides to break clumps apart
- 5. Filter mixed sample through tea strainer into clean cup. Push around to get all liquid and tap until sample goes into a ball
- 6. Use pipettes to fill 2 chambers of the slide with ~1ml sample
- 7. Let sit 5 mins for eggs to rise to stop
- 8. Place under slide and count strongyle eggs in each chamber following the grid lines on 10x power
- 9. Multiply number of eggs in each chamber by 25 to get estimated total





Strongyle eggs - oval-shaped light brown center Ascarid eggs – can be counted but strongyles should be the focus If not sure which it is – count it



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